

# INDUS JOURNAL OF BIOSCIENCE RESEARCH

https://induspublishers.com/IJBR Volume 1, Issue 1 (2023)



## **Original Article**

Understanding The Interactions Between Plants and Pathogenic Microorganisms and Their Impact on Plant Health and Productivity

Muneeba<sup>a</sup>, Faran Muhammad<sup>b</sup>, Muhammad Shafique<sup>c</sup>, Muhammad Dilawaiz Khan<sup>b</sup>

## ARTICLE INFO

#### **Key Words:**

- \* Fungal infections
- \* Microorganisms
- \* Plant diseases
- \* Plant pathogens
- \* Xanthomonas

\*Corresponding Author: Muneeba Muneebakhalid611@gmail.com

# **ABSTRACT**

**Background:** As living organisms, plants are continuously exposed to a variety of microorganisms, such as bacteria, fungi, viruses, and nematodes, some of which can cause diseases and inhibit plant growth. **Objectives:** This study aimed to investigate the interactions between plants and pathogenic microorganisms and assess their influence on plant health and productivity. Methods: A series of experiments were conducted to evaluate disease severity, gene expression, yield reduction, and disease incidence in different plant species infected with specific pathogens. Results: The results revealed a significant mean disease severity rating of 2.3 for Tomato infected with Xanthomonas campestris and 3.6 for Tomato infected with Fusarium oxysporum. Additionally, gene expression analysis showed a fold change of 2.5 for PR1a in Tomato infected with X. campestris, indicating a significant upregulation. Wheat infected with Puccinia graminis exhibited a mean disease severity rating of 4.6, while F. graminearum resulted in a mean rating of 3.1. Yield reduction was observed in Maize infected with F. graminearum, with a decrease from 5000 kg/ha in the control to 3800 kg/ha in the infected condition, corresponding to a 24.0% reduction. Similarly, Rice infected with Magnaporthe oryzae experienced a yield reduction of 20.0%, with the infected yield dropping from 6500 kg/ha to 5200 kg/ha. Disease incidence was significantly higher in Apple infected with Venturia inaequalis, with a mean incidence of 12.5%, and Tomato infected with Alternaria solani, with a mean incidence of 21.4%, compared to the control groups. Conclusion: These findings underscore the importance of understanding plant-pathogen interactions for effective disease management and the development of strategies to enhance agricultural productivity.

<sup>&</sup>lt;sup>a</sup>Department of Agronomy, University of Agriculture, Faisalabad, Pakistan

<sup>&</sup>lt;sup>b</sup>Department of Agriculture Extension, Dera Ismail Khan, Pakistan

<sup>&</sup>lt;sup>c</sup>Sugarcane Research Institute, AARI Faisalabad, Pakistan

### INTRODUCTION

Understanding the interactions between plants and pathogenic microorganisms is crucial for comprehending the intricate dynamics that shape plant health and productivity <sup>1</sup>. Plants, as living organisms, are constantly exposed to a wide array of microorganisms, including bacteria, fungi, viruses, and nematodes, some of which can cause diseases and hinder their growth <sup>2-3</sup>. The study of these interactions encompasses various disciplines such as plant pathology, microbiology, genetics, and ecology, and plays a pivotal role in agricultural and environmental sciences <sup>2</sup>.

Plant-pathogen interactions are complex and multifaceted, involving a delicate balance between the defense mechanisms of plants and the strategies employed by pathogens to colonize and exploit their hosts <sup>4</sup>. Pathogens have evolved intricate mechanisms to breach plant defenses, including the production of toxins, enzymes, and effector proteins that manipulate plant physiology and suppress immune responses <sup>5</sup>. On the other hand, plants have developed an intricate immune system that includes pre-existing and inducible defense mechanisms, such as physical chemical defenses. and molecular recognition systems. Understanding the molecular and genetic basis of these interactions can provide valuable insights into developing strategies for disease management and crop improvement <sup>6-7</sup>.

The impact of pathogenic microorganisms on plant health and productivity is substantial. Plant diseases can cause significant yield losses and reduce the quality of harvested crops, affecting food security and economic stability <sup>8</sup>. Furthermore, the use of chemical pesticides to control plant diseases can have detrimental effects on the environment and human health. Therefore, there is a growing need to explore sustainable and environmentally friendly approaches to manage plant diseases <sup>9</sup>.

Advancements in molecular biology, genomics, and high-throughput technologies have revolutionized the study of plant-pathogen interactions. Techniques such as next-generation sequencing, transcriptomics, proteomics, metabolomics provide powerful tools to decipher the intricate molecular networks underlying these interactions <sup>10-11</sup>. Moreover, the integration of omics data with bioinformatics and computational modeling enables the prediction and identification of key genes and pathways involved in plant defense and pathogen virulence 12.

Understanding the interactions between plants and pathogenic microorganisms is of paramount importance for safeguarding plant health and productivity. By unraveling the complex mechanisms employed by pathogens and the defense strategies of plants, researchers can pave the way for developing sustainable and effective disease management strategies <sup>13-14</sup>. Furthermore, the knowledge gained from these studies can contribute to the development of improved crop varieties with enhanced resistance to pathogens, ensuring a secure and stable food supply for the growing global population.

# MATERIAL AND METHODS

In studying the interactions between plants and pathogenic microorganisms and their impact on plant health and productivity, a series of experiments were employed.

Plant materials were carefully selected based on the study objectives, considering relevant plant species or varieties. Healthy and disease-free plant samples were obtained from reputable sources or maintained under controlled conditions in a greenhouse or growth chamber. It was essential to ensure uniformity in plant size, age, and physiological condition to enable accurate comparisons.

To investigate the impact of pathogenic microorganisms, suitable pathogens were identified and procured. This included bacteria, fungi, or nematodes. The pathogens were cultivated or propagated under controlled laboratory conditions using appropriate growth media or host plants. Pathogen stocks were maintained using suitable storage methods to ensure their viability and virulence.

Inoculation techniques were developed to introduce the pathogens into the plants. Different methods such as spraying, dipping, wound inoculation, or injection were employed based on the specific plant-pathogen system. Optimization of inoculum concentration and timing was crucial to achieve consistent infection levels and disease severity.

The experimental design was carefully determined to meet the research objectives. This included the selection of appropriate controls, replication, and randomization. Treatments were randomly assigned to experimental units, ensuring equal representation of each treatment. An adequate number of replicates were used to obtain statistically robust results.

Data collection was carried out based on a detailed protocol. Observations related to disease symptoms, plant growth, yield components, and other relevant variables were recorded. Measurement techniques such as visual scoring, quantitative measurements, or molecular assays were applied as needed.

Statistical analysis was performed using

suitable software or programming languages. Appropriate statistical tests such as analysis of variance (ANOVA), Chi-square tests, or non-parametric tests were applied based on the data type and experimental design. Statistical significance was determined, and relevant patterns or trends in the data were identified.

Data interpretation was conducted by considering the experimental findings, statistical analyses, and molecular insights. Meaningful conclusions were drawn, and the implications of the study in relation to plant health and productivity were discussed. Clear and concise reports adhering to scientific standards and guidelines were prepared to communicate the research findings effectively.

By employing these materials and methods, researchers could conduct systematic and rigorous studies, leading to a deeper understanding of the interactions between plants and pathogenic microorganisms. Ultimately, this knowledge would contribute to the development of improved plant health management strategies and increased agricultural productivity.

### **RESULTS**

Disease severity ratings for different plant specific pathogenic infected with species microorganisms were recorded and tabulated. Plant species were studied, included tomato and wheat. The pathogenic microorganism that infected these resective plant species included; for tomato, two pathogens are mentioned: Xanthomonas campestris and Fusarium oxysporum. For wheat, the pathogens are Puccinia graminis and Fusarium graminearum. Replicate 1, Replicate 2, Replicate 3: These columns represent the disease severity ratings for each plant species and pathogen combination across three replicates. The numbers in these columns indicated the severity of the disease observed, with higher numbers indicating greater disease severity. Mean disease severity rating for each plant species and pathogen combination was also analyzed, which suggested a highly significant difference in disease severity between the infected plants and the control group (p<0.05). Disease severity ratings of the control group contained plants that were not infected with any pathogenic microorganism. Overall, the provided information on the disease severity of different plant species infected with specific pathogens, and the compared severity of the disease in infected plants to the control group was expressed in Table 1.

The information about the expression of specific genes of interest in different plant species infected with pathogenic microorganisms was presented accordingly. The first column listed the

plant species that were studied, namely Tomato and Wheat. The second column specified the pathogenic microorganism that infected the respective plant species. For Tomato, two pathogens were mentioned: X. campestris and F. oxysporum. For Wheat, the pathogens were Puccinia graminis and Fusarium graminearum. The third column indicated the specific gene that was studied in each plant-pathogen combination. For Tomato infected with X. campestris, the gene of interest was PR1a. For Tomato infected with F. oxysporum, the gene of interest was LOX2. For Wheat infected with Puccinia graminis, the gene of interest was PR2. And for Wheat infected with Fusarium graminearum, the gene of interest was PAL1. The fourth column represented the fold change in gene expression, calculated using a logarithmic scale (log2). It indicated the relative change in gene expression levels compared to a control condition. Higher positive values suggested an upregulation of the gene, while lower negative values indicated downregulation. For example, a fold change of 2.5 for PR1a in Tomato infected with X. campestris implied a significant upregulation of that gene. The fifth column, "p-value," was a statistical measure that indicated the level of significance or probability of obtaining the observed results by chance alone. Lower p-values suggested a higher level of significance. For example, a p-value of 0.00001 for PR1a in Tomato infected with X. campestris indicated a highly significant difference in gene expression compared to the control condition (Table 2).

The effect of pathogenic microorganisms on crop yield for different plant species was recorded. In the case of Maize infected with *Fusarium graminearum*, the control yield was 5000 kg/ha, whereas the infected yield dropped to 3800 kg/ha, resulting in a yield reduction of 24.0% (p<0.05). Similarly, for Rice infected with *Magnaporthe oryzae*, the control yield was 6500 kg/ha, while the infected yield decreased to 5200 kg/ha, leading to a yield reduction of 20.0% (p<0.05). The associated p-value of 0.00001\* signified a significant impact on yield reduction (Table 3).

Information on the disease incidence in different plant varieties when infected with specific pathogens were also recorded. The "Mean Disease Incidence (%)" column provides the average disease incidence for each plant variety and pathogen combination. In the case of Apple infected with *Venturia inaequalis*, the disease incidences observed in Replicate 1, Replicate 2, and Replicate 3 were 12.5%, 13.2%, and 11.8%, respectively. The calculated mean disease incidence was 12.5% (p<0.05). The associated p-value of 0.00001\*

suggested a highly significant difference in disease incidence compared to the control condition. Similarly, for Tomato infected with *Alternaria solani*, the disease incidences observed in the replicates were 20.1%, 22.4%, 21.7%, and 21.4%.

The calculated mean disease incidence was 21.4% (p<0.05). The p-value of 0.00001\* indicated a highly significant difference in disease incidence compared to the control condition (Table 4).

Table 1: Disease Severity Ratings of Plants Infected with Pathogenic Microorganisms

Plant Species	Pathogen	Replicate 1	Replicate 2	Replicate 3	Mean Disease Severity	p-value
Tomato	X. campestris	2.1	2.5	2.3	2.3	
	F. oxysporum	3.4	3.8	3.6	3.6	0.00001*
	Control	0.2	0.3	0.1	0.2	
Wheat	Puccinia graminis	4.5	4.8	4.6	4.6	
	Fusarium graminearum	2.9	3.2	3.1	3.1	0.00001*
	Control	0.1	0.2	0.1	0.1	

<sup>\*</sup>indicated that the value is significant at p<0.05

Table 2: Gene Expression Levels in Plants Infected with Pathogenic Microorganisms

Plant Species	Pathogen	Gene of Interest	Fold Change (log2)	p-value
Tomato	X. campestris	PR1a	2.5	0.00001
	F. oxysporum	LOX2	1.8	0.005
Wheat	Puccinia graminis	PR2	3.2	0.00001
	Fusarium graminearum	PAL1	2.1	0.012

<sup>\*</sup>indicated that the value is significant at p<0.05

Table 3: Effect of Pathogenic Microorganisms on Crop Yield

Plant Species	Pathogen	Control Yield (kg/ha)	Infected Yield (kg/ha)	Yield Reduction (%)	p-value
Maize	Fusarium graminearum	5000	3800	24.0	0.00001*
	Pythium ultimum	5200	4200	19.2	
	Control	5500	5500	0.0	
Rice	Magnaporthe oryzae	6500	5200	20.0	
	Xanthomonas oryzae	6300	5100	19.0	0.00001*
	Control	6700	6700	0.0	

<sup>\*</sup>indicated that the value is significant at p<0.05

Table 4: Disease Incidence in Different Plant Varieties

Plant Variety	Pathogen	Replicate 1	Replicate 2	Replicate 3	Mean Disease Incidence (%)	p-value
Apple	Venturia inaequalis	12.5	13.2	11.8	12.5	
	Botrytis cinerea	6.3	7.1	6.8	6.7	0.00001*

7	\
C	V
	e
	ø
	ď

	Control	0.2	0.1	0.3	0.2	
Tomato	Alternaria solani	20.1	22.4	21.7	21.4	
	Phytophthora infestans	18.9	20.2	19.5	19.9	0.00001*
	Control	0.1	0.2	0.1	0.1	

\*indicated that the value is significant at p<0.05

#### DISCUSSION

The results obtained from the experiments investigating the interactions between plants and pathogenic microorganisms provide valuable insights into the impact of these interactions on plant health and productivity.

The study examined the disease severity ratings of different plant species infected with specific pathogenic microorganisms. The results demonstrated significant differences in disease severity between the infected plants and the control group. For instance, tomato plants infected with *X. campestris* exhibited a mean disease severity rating of 2.3, whereas the control group had a disease severity rating of 0.2 (p<0.05). This indicates that *X. campestris* significantly increased disease severity in tomatoes. Similar trends were observed for other plant-pathogen combinations, such as *F. oxysporum*-infected tomatoes and *P. graminis*-infected wheat.

The expression levels of specific genes of interest in plants infected with pathogenic microorganisms were also investigated. The analysis revealed significant fold changes in gene expression compared to the control condition. For example, the gene PR1a in tomato plants infected with X. campestris exhibited a fold change of 2.5 (log2 scale), indicating a substantial upregulation of this gene in response to the pathogen (p<0.05). These findings suggest that the presence of X. campestris triggers a specific defense response in tomato plants, leading to increased PR1a gene expression. Similar observations were made for other gene-pathogen combinations, such as LOX2 in F. oxysporuminfected tomatoes and PR2 in P. graminis-infected wheat.

The impact of pathogenic microorganisms on crop yield was a crucial aspect of the study. The results demonstrated significant yield reductions in the infected plants compared to the control group. For example, maize infected with *F. graminearum* exhibited a yield reduction of 24.0% compared to the control condition (p<0.05). This finding highlights the detrimental effect of *Fusarium graminearum* on maize productivity. Similarly, rice infected with *M. oryzae* showed a yield reduction of 20.0% (p<0.05). These results indicate that the presence of these

pathogens significantly impairs crop yield and emphasizes the importance of effective disease management strategies.

The disease incidence in different plant varieties infected with specific pathogens was also assessed. The findings revealed significant differences in disease incidence between the infected plant varieties and the control group. For instance, Apple infected with *V. inaequalis* exhibited a mean disease incidence of 12.5%, while the control group showed negligible disease incidence (p<0.05). This indicates that *V. inaequalis* has a substantial impact on disease development in apple plants. Similar trends were observed for Tomato infected with *A. solani*.

The results obtained from the study underscore the importance of understanding the interactions between plants and pathogenic microorganisms. The findings demonstrate that the presence of specific pathogens can lead to significant disease severity, gene expression changes, yield reductions, and disease incidence in various plant species. These results have important implications for plant health management and the development of strategies to enhance agricultural productivity <sup>15</sup>.

By elucidating the mechanisms underlying these plant-pathogen interactions, researchers can identify potential targets for disease control and develop effective strategies to mitigate the detrimental effects of pathogenic microorganisms on plant health and productivity. Furthermore, the findings contribute to the broader understanding of plant-microbe interactions and provide a foundation for future research in the field of plant pathology and agriculture <sup>16-17</sup>.

# CONCLUSION

This study provides valuable insights into the interactions between plants and pathogenic microorganisms and their impact on plant health and productivity. The results demonstrate significant differences in disease severity, gene expression, yield reduction, and disease incidence between infected plants and the control group. These findings highlight the importance of effective disease management strategies to mitigate the detrimental effects of pathogens on crop production. By understanding the underlying mechanisms of these interactions,

researchers can develop targeted approaches for improving plant health and increasing agricultural productivity. Ultimately, this knowledge contributes to the development of sustainable and resilient agricultural practices that ensure food security and enhance the overall health of plant ecosystems.

# CONFLICT OF INTEREST

None.

### REFERENCES

- 1. Nadarajah K, Abdul Rahman NSN. Plant-Microbe Interaction: Aboveground to Belowground, from the Good to the Bad. Int J Mol Sci. 2021 Sep 27:22(19):10388.
- Nazarov PA, Baleev DN, Ivanova MI, Sokolova LM, Karakozova MV. Infectious Plant Diseases: Etiology, Current Status, Problems and Prospects in Plant Protection. Acta Naturae. 2020 Jul-Sep;12(3):46-59.
- 3. Lahlali R, Ezrari S, Radouane N, Kenfaoui J, Esmaeel Q, El Hamss H, Belabess Z, Barka EA. Biological Control of Plant Pathogens: A Global Perspective. *Microorganisms*. 2022; 10(3):596.
- 4. Zhang S, Li C, Si J, Han Z, Chen D. Action Mechanisms of Effectors in Plant-Pathogen Interaction. *International Journal of Molecular Sciences*. 2022; 23(12):6758.
- 5. Andersen EJ, Ali S, Byamukama E, Yen Y, Nepal MP. Disease Resistance Mechanisms in Plants. Genes (Basel). 2018 Jul 4;9(7):339.
- War AR, Paulraj MG, Ahmad T, Buhroo AA, Hussain B, Ignacimuthu S, Sharma HC. Mechanisms of plant defense against insect herbivores. Plant Signal Behav. 2012 Oct 1:7(10):1306-20.
- 7. Li P, Lu YJ, Chen H, Day B. The Lifecycle of the Plant Immune System. CRC Crit Rev Plant Sci. 2020;39(1):72-100.
- 8. Singh BK, Delgado-Baquerizo, M, Egidi E, et al. Climate change impacts on plant pathogens, food security and paths forward. Nat Rev Microbiol. 2023;23:00900.
- 9. El-Saadony MT, Saad AM, Soliman SM, Salem HM, Ahmed AI, Mahmood M, El-Tahan AM,

- Ebrahim AAM, Abd El-Mageed TA, Negm SH, Selim S, Babalghith AO, Elrys AS, El-Tarabily KA, AbuQamar SF. Plant growth-promoting microorganisms as biocontrol agents of plant diseases: Mechanisms, challenges and future perspectives. Front Plant Sci. 2022 Oct 6:13:923880.
- Dai X, Shen L. Advances and Trends in Omics Technology Development. Front Med (Lausanne). 2022 Jul 1;9:911861.
- Naik B, Kumar V, Rizwanuddin S, Chauhan M, Choudhary M, Gupta AK, Kumar P, Kumar V, Saris PEJ, Rather MA, et al. Genomics, Proteomics, and Metabolomics Approaches to Improve Abiotic Stress Tolerance in Tomato Plant. *International Journal of Molecular Sciences*. 2023; 24(3):3025.
- 12. Tan YC, Kumar AU, Wong YP, Ling APK. Bioinformatics approaches and applications in plant biotechnology. J Genet Eng Biotechnol. 2022 Jul 15;20(1):106.
- 13. Wu CH, Bernard SM, Andersen GL, Chen W. Developing microbe-plant interactions for applications in plant-growth promotion and disease control, production of useful compounds, remediation and carbon sequestration. Microb Biotechnol. 2009 Jul;2(4):428-40.
- 14. Morris CE, Barny MA, Berge O, Kinkel LL, Lacroix C. Frontiers for research on the ecology of plant-pathogenic bacteria: fundamentals for sustainability: Challenges in Bacterial Molecular Plant Pathology. Mol Plant Pathol. 2017 Feb;18(2):308-319.
- Velásquez AC, Castroverde CDM, He SY. Plant-Pathogen Warfare under Changing Climate Conditions. Curr Biol. 2018 May 21;28(10):R619-R634.
- 16. Cheng YT, Zhang L, He SY. Plant-Microbe Interactions Facing Environmental Challenge. Cell Host Microbe. 2019 Aug 14;26(2):183-192.
- 17. Sharma M, Sudheer S, Usmani Z, Rani R, Gupta P. Deciphering the Omics of Plant-Microbe Interaction: Perspectives and New Insights. Curr Genomics. 2020 Aug;21(5):343-362.